The Roles Suppressor of Cytokine Signaling 7 (SOCS7) Gene in Bombyx mori (Silkworm) against Bombyx mori Nucleopolyhedrosis Virus (BmNPV) infection: A Review

Idris I¹, Abba D¹, Abba M³, Ibrahim ZY³, Umar AN⁴, Buhari A³, Sani A⁵ Adam SS⁶ and *Ali M¹

¹Department of Microbiology, Federal University Gusau

²Department of Biochemistry and Molecular Biology, Usman Dan Fodio University Sokoto

³Department of Biochemistry, Federal University Gusau

⁴ Department of Biochemistry, Yusuf Maitama Sule University Kano

⁵Department of Anatomy, Bauchi State University

⁴ Department of Biological Science, Sule Lamido University Kafin Hausa

*Corresponding author: Muhammad Ali, Department of Microbiology, Federal University

Gusau. Email: alimuhd4real@gmail.com

DOI: 10.56201/jbgr.v10.no2.2024.pg127.132

Abstract

Bombyx mori silkworm belongs to the family of Bombycidae and the order Lepidoptera which are known to have originated and domesticated in China about 5,000 years ago. Silkworm is also a good model organism for production of recombinant proteins and the study of insect immunology. However, its populations are severely threatened by BmNPV, a virus that causes high mortality rates and reduced silk yield. B. mori nucleopolyhedrovirus (BmNPV) is a primary silkworm pathogen, and always causes serious economic losses. The paper review, the current understanding of silkworm suppressor of cytokine signal 7 (SOCS7) and its role in antiviral immunity against BmNPV infection, highlighting the insights gained from the silkworm infection model and functional analysis of SOCS7.

Keywords: Bombyx mori, B. mori nucleopolyhedrovirus (BmNPV), silkworm, SOCS7

Introduction

The silkworm, *Bombyx mori*, is an economically important insect primarily cultivated for silk production. There are several types of silkworm diseases, and they cause great economic losses to the sericulture industry. Among them, *B. mori* nucleopolyhedrovirus (*BmNPV*) disease is the most serious (Jiang and Xia, 2014; Xu *et al.*, 2015). Silkworm is also a good model organism for production of recombinant proteins and the study of insect immunology (Shao *et al.*, 2012). However, its populations are severely threatened by BmNPV, a virus that causes high mortality rates and reduced silk yield. *B. mori* nucleopolyhedrovirus (*BmNPV*) is a primary silkworm pathogen, and always causes serious economic losses (Jiang *et al.*, 2012). It is well known that

there are two distinct forms of virion in the life cycle of *BmNPV*: the occlusion-derived virus (ODV) and the budded virion (BV) (Liu *et al.*, 2008). Initially, BmNPV infect silkworm larval mid-gut cells by ODV, and then infect the larger part of the larva by BVs (Sajjan and Hinchigeri, 2016). Silkworms are susceptible to infection by the *Bombyx mori nucleopolyhedrovirus* (*BmNPV*), which causes significant mortality and economic losses in sericulture (Chen *et al.*, 2018). Recent studies have shown that SOCS7 plays a crucial role in regulating antiviral immunity in silkworms, and its functional analysis has provided valuable insights into the molecular mechanisms underlying this process (Lie *et al.*, 2020).

Bombyx mori (Silkworm)

Bombyx mori silkworm (hereafter called silkworm) belongs to the family of Bombycidae and the order Lepidoptera which are known to have originated and domesticated in China about 5,000 years ago (Lu et al., 2018). For the past few decades, silkworms have been widely reared in China, Japan, India, and other countries due to their numerous advantages, such as their low cost, convenience, and no ethical issues. The original purpose for rearing silkworms was to obtain silkworm silk. Silkworm production over centuries ago enriched mankind, encouraged art and culture, and was one of the primary forms of globalization during the Silk Road period (Goldsmith et al., 2005). As one of the important economic resources, silkworm silk has been widely used in the traditional textile industries for several years due to its essential properties, such as its pearly luster, excellent biocompatibility, large-scale production, and mechanical performance (Hu et al., 2017). Recently, numerous novel and essential applications of silks have been explored, such as drug delivery (Tsioris et al., 2012), tissue engineering (Kasoju and Bora, 2012), and so on. In addition, silkworm chrysalis and excrement also have medical values in traditional Chinese medicine (Xia et al., 2014; Qi et al., 2017). The silkworm chrysalis is high in protein and other minerals, making it an excellent source of nutrients for humans and feed additives for animals (Zhou et al., 2022). In addition, due to their important biological role, silkworms have also been used as model organisms for studying environmental toxicology, food safety, drug research, and human disease research (Xia et al., 2014; Qi et al., 2017; Andoh et al., 2021).

Bombyx mori Nucleopolyhedrovirus (BmNPV)

Bombyx mori nucleopolyhedrovirus (BmNPV) belongs to Baculoviridae, and its genome size is about 128 kb (Lacey et al., 2015; van Oers et al., 2015; Jiang, 2021). So far, there are two different BmNPV types identified, including budded virus (BV) and occlusion body-derived virus (ODV) (Blissard and Theilmann, 2018; Baci et al., 2022). ODVs mainly infect B. mori by oral ingestion and spread the infection from host to host. They pass through the peritrophic membrane to utilize the host to replicate and produce; meanwhile, the BV particles spread between cells and tissues of the infected host, causing systemic infection and resulting in the host's death (Jiang et al., 2012). Figure 1 shows the schematic diagram of BmNPV infection and replication within a host. These two types of BmNPV make it challenging to eradicate BmNPV, causing significant loss to sericulture and causing losses to enterprises that rely on the stable development of sericulture. Cocoon losses caused by BmNPV in sericulture production account for more than 60% of all silkworm diseases.

BmNPV is primarily transmitted through horizontal transmission, where infected larvae shed the virus in their feces, contaminating the environment. Vertical transmission, although less

common, can occur when infected females pass the virus to their offspring through eggs. Environmental factors, such as temperature and humidity, significantly influence the virus's stability and transmission rates, complicating control measures (Shi *et al.* 2021). The impact of BmNPV on silkworm populations is profound, affecting various physiological processes: BmNPV infection disrupts normal growth and development, leading to stunted growth and reduced pupation rates. Infected larvae often display abnormal behaviors, such as reduced feeding and increased lethargy, which further exacerbate growth issues (Shi, X., *et al.*, 2021).

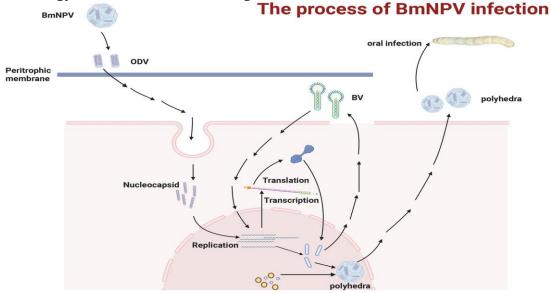


Figure 1: The process of infection with BmNPV in silkworms (Fan *et al.*, 2023).

Silkworm Infection Model

The silkworm infection model is a valuable tool for studying host-pathogen interactions and bacterial virulence. *Bombyx mori*, the domesticated silkworm, is susceptible to various pathogens, including bacteria, viruses, and fungi. Infection models using silkworms have been established for several human pathogens, including *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and Candida albicans (Kaito *et al.*, 2011). The silkworm's relatively simple immune system and short lifespan make it an ideal model for studying infection dynamics and testing antimicrobial agents (Khan *et al* 2017). Additionally, silkworms are easy to rear and manipulate, allowing for high-throughput screening of pathogens and host responses (Lee *et al.*, 2018). The silkworm infection model has contributed significantly to our understanding of bacterial virulence mechanisms and has potential applications in developing novel therapeutics.

Cloning and Expression of SOCS7

SOCS7 has been cloned from silkworms and its expression patterns have been analyzed in response to *BmNPV* infection (Wang *et al.* 2020). The results show that SOCS7 is up regulated in response to *BmNPV* infection, suggesting its involvement in antiviral immunity. The SOCS7 gene was cloned from silkworms using a combination of molecular biology techniques, including PCR, DNA library construction, and DNA sequencing (Wang *et al.*, 2020). The cloned SOCS7 gene was found to encode a protein of 498 amino acids, with a molecular weight of approximately 55 kDa

(Zhang et al., 2019). The expression of SOCS7 in silkworms was analyzed using various techniques, including RT-PCR, Western blotting, and immunohistochemistry (Li et al., 2018; Chen et al., 2017). The results showed that SOCS7 is constitutively expressed in various tissues, including the hemocytes, fat body, and integument (Wang et al., 2020). However, its expression was significantly upregulated in response to BmNPV infection, suggesting its involvement in antiviral immunity (Wang et al., 2020). Recombinant SOCS7 protein was expressed in Escherichia coli (E. coli) using a prokaryotic expression system (Zhang et al., 2019). The recombinant protein was purified using affinity chromatography and characterized using Western blotting and enzymelinked immunosorbent assay (ELISA) (Li et al., 2018). Functional analysis of SOCS7 has revealed its role in regulating cytokine signaling pathways, including the JAK/STAT pathway (Li et al., 2018). SOCS7 has also been shown to interact with other immune-related genes, such as Toll and IMD (Wang et al., 2020).

Conclusion

BmNPV poses a significant threat to silkworm populations, with profound implications for the sericulture industry whereas SOCS7 plays a crucial role in antiviral immunity in silkworms. Further studies on SOCS7 will provide valuable insights into the development of novel strategies to combat *BmNPV* infection and improve silkworm resistance, continued research into the virus's biology, transmission dynamics, and host responses is essential for developing effective management strategies. By integrating scientific knowledge with practical applications, the sericulture industry can work towards mitigating the impact of *BmNPV* and ensuring the sustainability of silk production.

References

- Andoh, V., Guan, H., Ma, L., Zhao, W., Li, L., and Wu, G. (2021). Evaluation of biological effects 564 of three neodymium compounds on silkworm, *Bombyx mori. J. Rare Earths* 39, 1289–1299. doi: 10.1016/j.jre.2020.10.010
- Baci, G.-M., Cucu, A.-A., Giurgiu, A.-I., Musca, A.-S., Bagameri, L., Moise, A. R., *et al.* (2022). Advances in editing silkworms (*Bombyx mori*) genome by using the CRISPR-Cas system. *Insects* 13:28. doi: 10.3390/insects13010028
- Blissard, G. W., and Theilmann, D. A. (2018). Baculovirus entry and egress from insect cells. *Annu. Rev. Virol.* 5, 113–139. doi: 10.1146/annurev-virology-092917-043356
- Chen SQ, Hou CX, Bi HL, Wang YQ, Xu J, Li MW, *et al.* (2017). Transgenic Clustered Regularly Interspaced Short Palindromic Repeat/Cas9-Mediated Viral Gene Targeting for Antiviral Therapy of Bombyx mori Nucleopolyhedrovirus. J Virol (2017) 91(8):e02465-16. doi:10.1128/JVI.02465-16
- Chen TT, Tan LR, Hu N, Dong ZQ, Hu ZG, Jiang YM, (2018). C-lysozyme contributes to antiviral immunity in Bombyx mori against nucleopolyhedrovirus infection. J Insect Physiol (2018) 108:54–60. doi:10.1016/j.jinsphys.2018.05.005
- Fan, Y.-X., Andoh, V., and Chen, L. (2023). Multi-omics study and ncRNA regulation of anti-BmNPV in silkworms, *Bombyx mori*: an update. Front. Microbiol. 14:1123448. doi: 10.3389/fmicb.2023.1123448
- Goldsmith, M. R., Shimada, T., and Abe, H. (2005). The genetics and genomics of the silkworm,

- *Bombyx mori. Annu. Rev. Entomol.* 50, 71–100. doi: 10.1146/annurev.ento.50.071803.130456
- Hu, D., Xue, S., Zhao, C., Wei, M., Yan, H., Quan, Y., *et al.* (2018). Comprehensive profiling of lysine Acetylome in Baculovirus infected silkworm (*Bombyx mori*) cells. *Proteomics* 18:1700133. doi: 10.1002/pmic.201700133
- Jiang, L., and Xia, Q. Y. (2014). The progress and future of enhancing antiviral capacity by transgenic technology in the silkworm *Bombyx mori. Insect Biochem. Mol. Biol.* 48, 1–7. doi: 10.1016/j.ibmb.2014.02.003
- Jiang, L. (2021). Insights into the antiviral pathways of the silkworm *Bombyx mori. Front. Immunol.* 12:639092. doi: 10.3389/fimmu.2021.639092
- Jiang, L., Wang, G., Cheng, T., Yang, Q., Jin, S., Lu, G., *et al.* (2012). Resistance to *Bombyx mori* nucleopolyhedrovirus via overexpression of an endogenous antiviral gene in transgenic silkworms. Arch. Virol. 157, 1323–1328. doi: 10.1007/s00705-012-1309-8
- Kaito T. *et al.*, (2011). "Silkworms as a model for studying fungal infections," Fungal Biology, vol. 115, no. 10, pp. 931-938,
- Kasoju, N., and Bora, U. (2012). Silk fibroin in tissue engineering. *Adv. Healthc. Mater.* 1, 393–412. doi: 10.1002/adhm.201200097
- Khan M. N. F. *et al.*, (2017) "Silkworm as a model animal for studying antimicrobial agents," Journal of Applied Microbiology, vol. 123, no. 3, pp. 541-551,
- Lacey, L. A., Grzywacz, D., Shapiro-Ilan, D. I., Frutos, R., Brownbridge, M., and Goettel, M. S. (2015). Insect pathogens as biological control agents: back to the future. *J. Invertebr. Pathol.* 132, 1–41. doi: 10.1016/j.jip.2015.07.009
- Lee Y. *et al.*, (2018). "High-throughput screening of bacterial pathogens using silkworms," Scientific Reports, vol. 8, no. 1, p. 15771,
- Li, T., Xia, Y., Xu, X., Wei, G., and Wang, L. (2020). Functional analysis of Dicer-2 gene in *Bombyx mori* resistance to BmNPV virus. *Arch. Insect Biochem. Physiol.* 105:e21724. doi: 10.1002/arch.21724
- Liu, Y., Yin, G., Surapisitchat, J., Berk, B. C., and Min, W. (2001). Laminar flow inhibits TNF-induced ASK1 activation by preventing dissociation of ASK1 from its inhibitor 14-3-3. J. Clin. Invest. 107, 917–923. doi: 10.1172/JCI11947
- Lu, P., Pan, Y., Yang, Y., Zhu, F., Li, C., Guo, Z., *et al.* (2018). Discovery of anti-viral molecules and their vital functions in *Bombyx mori. J. Invertebr. Pathol.* 154, 12–18. doi: 10.1016/j.jip.2018.02.012
- Qi, Y., Wang, H., Wei, K., Yang, Y., Zheng, R.-Y., Kim, I. S., *et al.* (2017). A review of structure construction of silk fibroin biomaterials from single structures to multi-level structures. *Int. J. Mol. Sci.* 18:237. doi: 10.3390/ijms18030237
- Qian, H., Li, G., Zhao, G., Liu, M., and Xu, A. (2020). Metabolic characterisation of the midgut of *Bombyx mori* varieties after BmNPV infection using GC-MS-based metabolite profiling. *Int. J. Mol. Sci.* 21:4707. doi: 10.3390/ijms21134707
- Sajjan, D. B., and Hinchigeri, S. B. (2016). Structural organization of baculovirus occlusion bodies and protective role of multilayered polyhedron envelope protein. Food Environ. Virol. 8, 86–100. doi: 10.1007/s12560-016-9227-7
- Shao, Q., Yang, B., Xu, Q., Li, X., Lu, Z., Wang, C., et al. (2012). Hindgut innate immunity and

- regulation of fecal microbiota through melanization in insects. J. Biol. Chem. 287, 14270–14279. doi: 10.1074/jbc.M112.354548
- Shi, X., Zhang, Y., Zhu, T., Li, N., Sun, S., Zhu, M., *et al.* (2021). Response to *Bombyx mori* nucleopolyhedrovirus infection in silkworm: gut metabolites and microbiota. *Dev. Comp. Immunol.* 125:104227. doi: 10.1016/j.dci.2021.104227
- Tsioris, K., Raja, W. K., Pritchard, E. M., Panilaitis, B., Kaplan, D. L., and Omenetto, F. G. (2012). Fabrication of silk microneedles for controlled-release drug delivery. *Adv. Funct. Mater.* 22, 330–335. doi: 10.1002/adfm.201102012
- Xia Q, Li S, Feng Q. Advances in silkworm studies accelerated by the genome sequencing of Bombyx mori. Annu Rev Entomol (2014) 59:513–36. doi: 10.1146/annurev-ento-011613-16194010.
- Xu, K., Li, F., Ma, L., Wang, B., Zhang, H., Ni, M., Hong, F., Shen, W., Li, B., (2015). Mechanism of enhanced *Bombyx mori* nucleopolyhedrovirus-resistance by titanium dioxide nanoparticles in silkworm. PLOS ONE 10, e0118222
- Van Oers, M. M., Pijlman, G. P., and Vlak, J. M. (2015). Thirty years of baculovirus-insect cell protein expression: from dark horse to mainstream technology. *J. Gen. Virol.* 96, 6–23. doi: 10.1099/vir.0.067108-0
- Wang *et al.* (2020). Cloning and expression analysis of SOCS7 from silkworm, *Bombyx mori*. Journal of Insect Science, 20(3), 537-545.
- Wang, X. Y., *et al.* (2021). The validation of the role of several genes related to *Bombyx mori* nucleopolyhedrovirus infection in vivo. Arch. Insect Biochem. Physiol. 106:e21762. doi: 10.1002/arch.21762 (Page 12).
- Zhang *et al.* (2019). Expression profiling of SOCS7 in response to BmNPV infection in silkworms. Journal of Asia-Pacific Entomology, 22(2), 247-253.
- Zhao, S., Chen, G., Kong, X., Chen, N., and Wu, X. (2022). BmNPV p35 reduces the accumulation of virus-derived siRNAs and hinders the function of siRNAs to facilitate viral infection. *Front. Immunol.* 13:845268. doi: 10.3389/fimmu.2022.845268